

Big Data Analytics and Molecular Diagnostic Methods for Treatment Pathways of Priority Pathogens



Webinar at ICGEB-DBT
Apex, BTIC, New Delhi

Anshu Bhardwaj, Ph.D.

Senior Scientist & Assistant Professor
Bioinformatics Center
CSIR-Institute of Microbial Technology
Chandigarh, India

WHO Priority Pathogens List (PPL) - 2017

CRITICAL	HIGH	MEDIUM
<i>Acinetobacter baumannii</i> , carbapenem-resistant	<i>Enterococcus faecium</i> , vancomycin-resistant	<i>Streptococcus pneumoniae</i> , penicillin-non-susceptible
<i>Pseudomonas aeruginosa</i> , carbapenem-resistant	<i>Staphylococcus aureus</i> , methicillin-resistant, vancomycin intermediate resistant	<i>Haemophilus influenzae</i> , ampicillin-resistant
<i>Enterobacteriaceae</i> *, carbapenem-resistant, 3 rd generation cephalosporin-resistant	<i>Neisseria gonorrhoeae</i> , 3 rd generation cephalosporin-resistant, fluoroquinolone-resistant	<i>Shigella</i> , fluoroquinolone-resistant
* Enterobacteriaceae include: <i>Klebsiella pneumoniae</i> , <i>Escherichia coli</i> , <i>Enterobacter spp.</i> , <i>Serratia spp.</i> , <i>Proteus spp.</i> , and <i>Providencia spp.</i> , <i>Morganella spp.</i>	<i>Campylobacter</i> , fluoroquinolone-resistant	ACCORDING TO THE STUDY, MTB IS ON THE PRIORITY FOR NEW DRUG DISCOVERY AND DEVELOPMENT
	<i>Salmonella</i> , fluoroquinolone-resistant	
	<i>Helicobacter pylori</i> , clarithromycin-resistant	

http://www.who.int/medicines/publications/WHO-PPL-Short_Summary_25Feb-ET_NM_WHO.pdf?ua=1



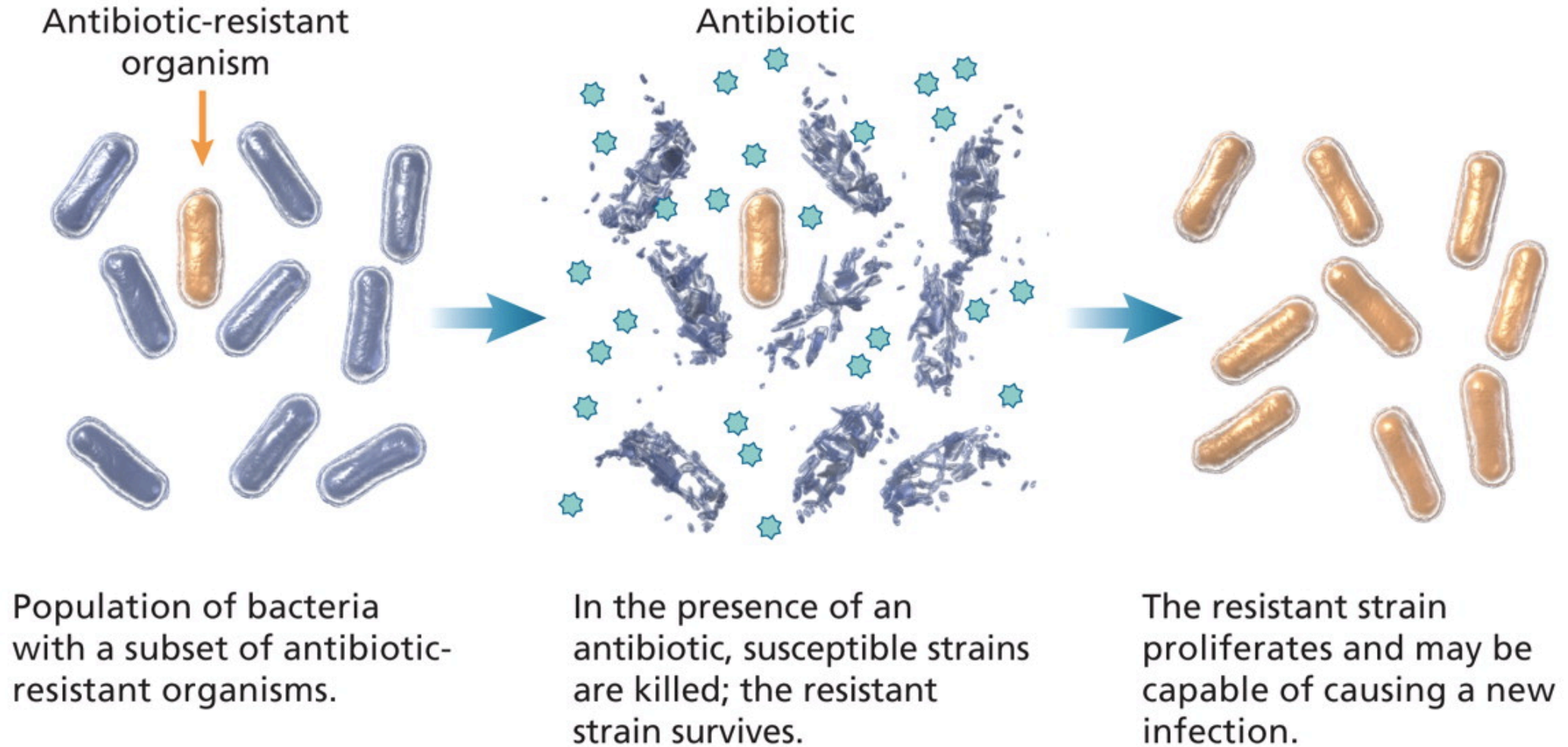
Antimicrobial resistance occurs when microorganisms such as bacteria, viruses, fungi and parasites change in ways that render the **medications** used to cure the infections they cause **ineffective**.

When the microorganisms become resistant to most antimicrobials they are often referred to as “**superbugs**”.



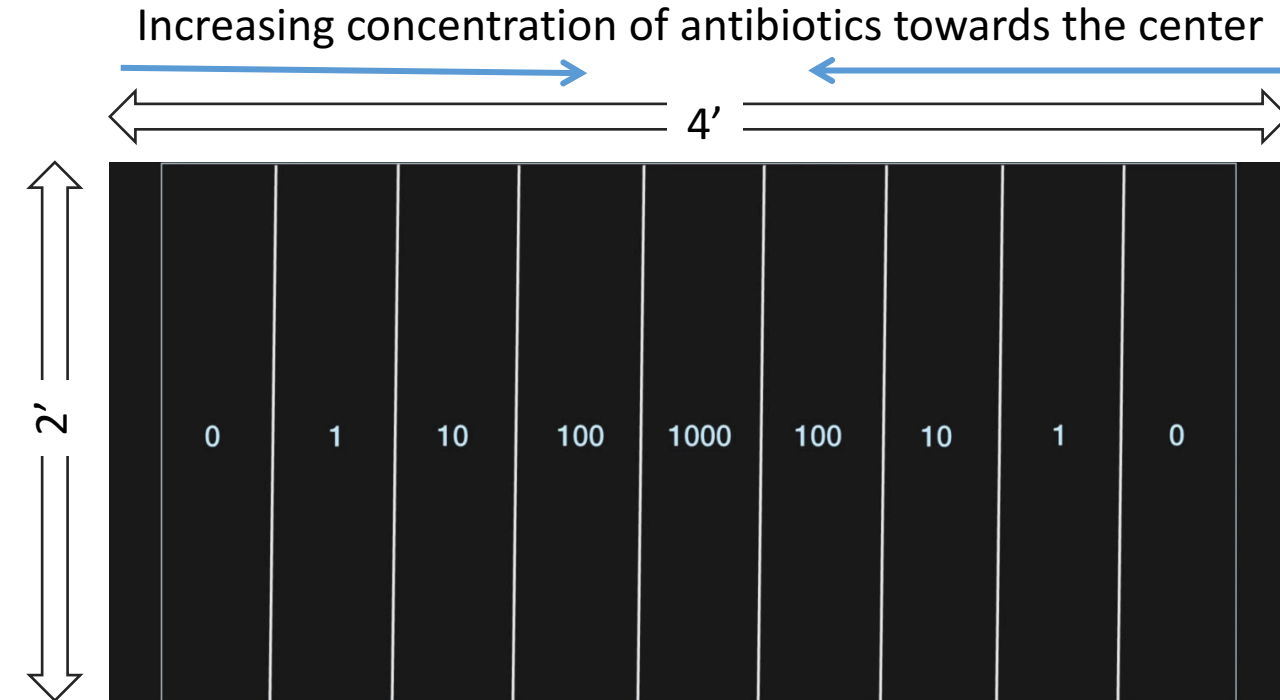
**World Health
Organization**

Antibiotic Resistance



<https://www.google.com/url?sa=i&source=imgres&cd=&cad=rja&uact=8&ved=2ahUKewifiOipQMDqAhXJxTgGHanwAPMQjRx6BAgBEAQ&url=https%3A%2F%2Fwww.cmaj.ca%2Fcontent%2F180%2F4%2F408%2Ftab-figures-data&psig=AOvVaw36qc4E5LvmsYs2UMdUTib2&ust=1594388842796509>

A cinematic approach to drug resistance – Harvard Gazette



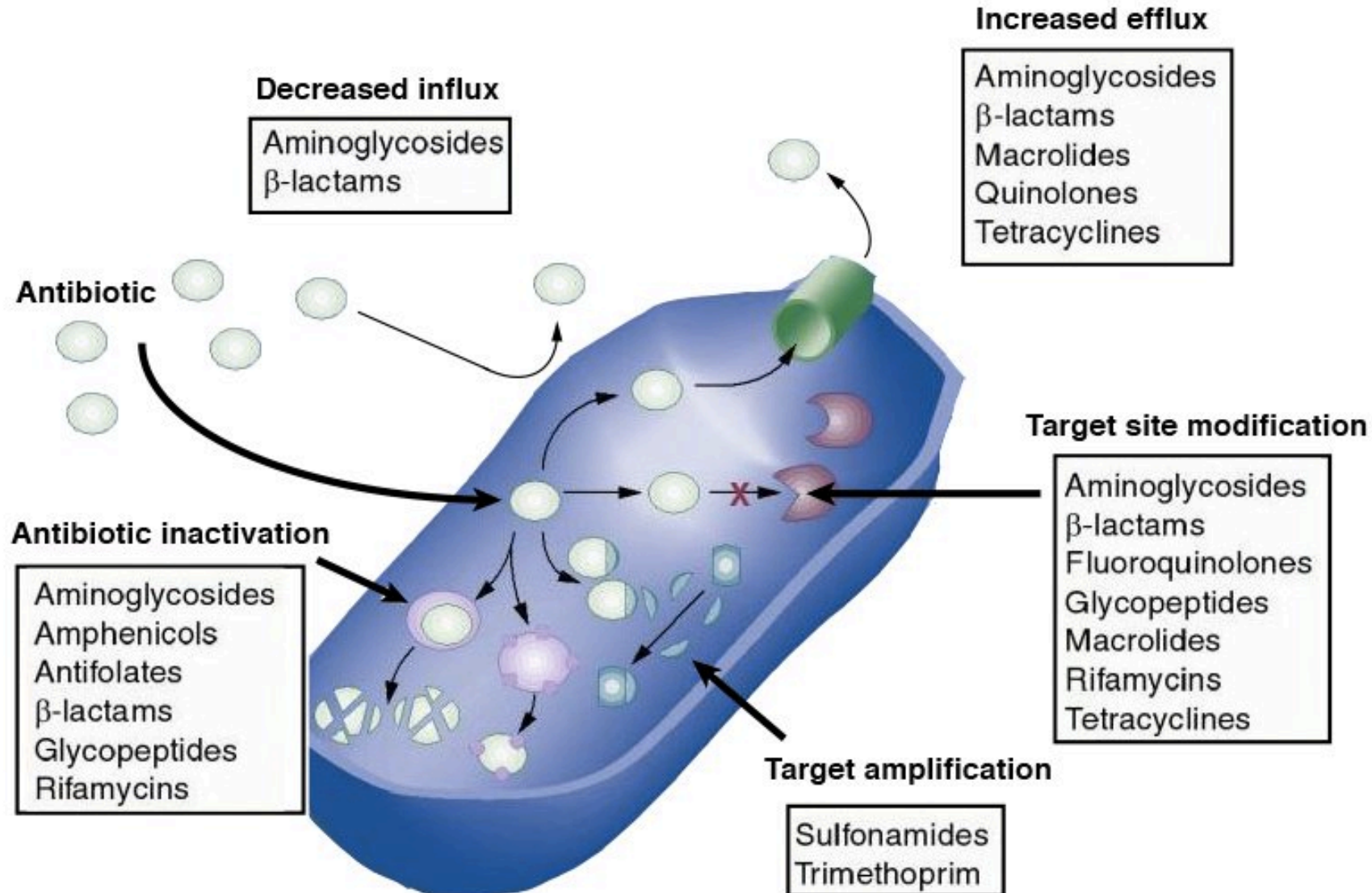
An experimental device for studying microbial evolution in a spatially structured environment

Microbial Evolution and Growth Arena (MEGA) plate



Bacteria spread as they evolve

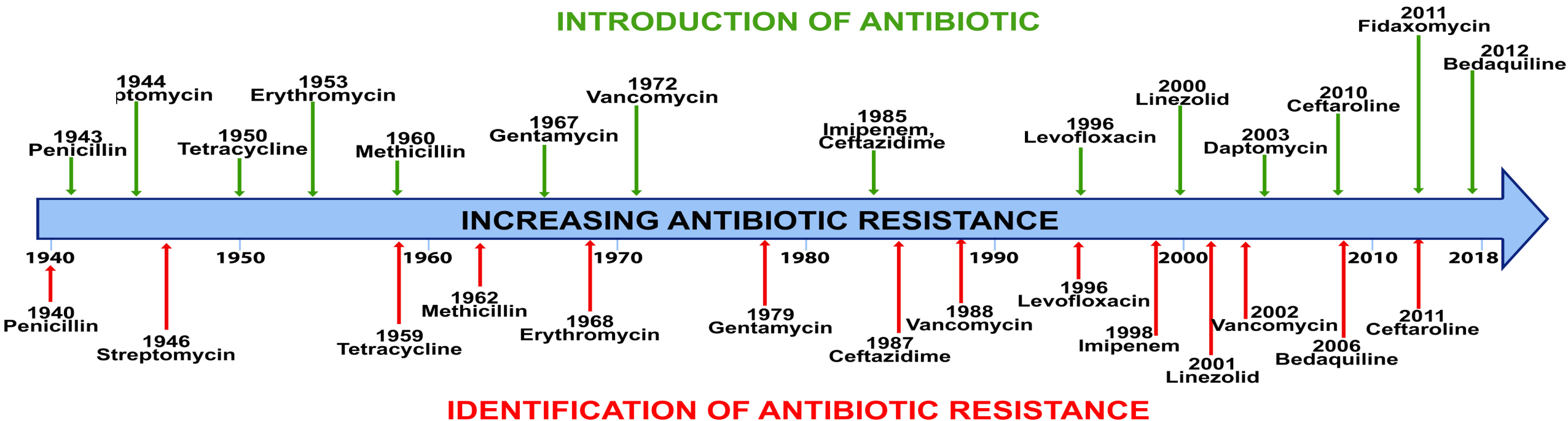
Drug Resistance Mechanisms



<https://adailydeed.wordpress.com/tag/antibiotic-resistance/>



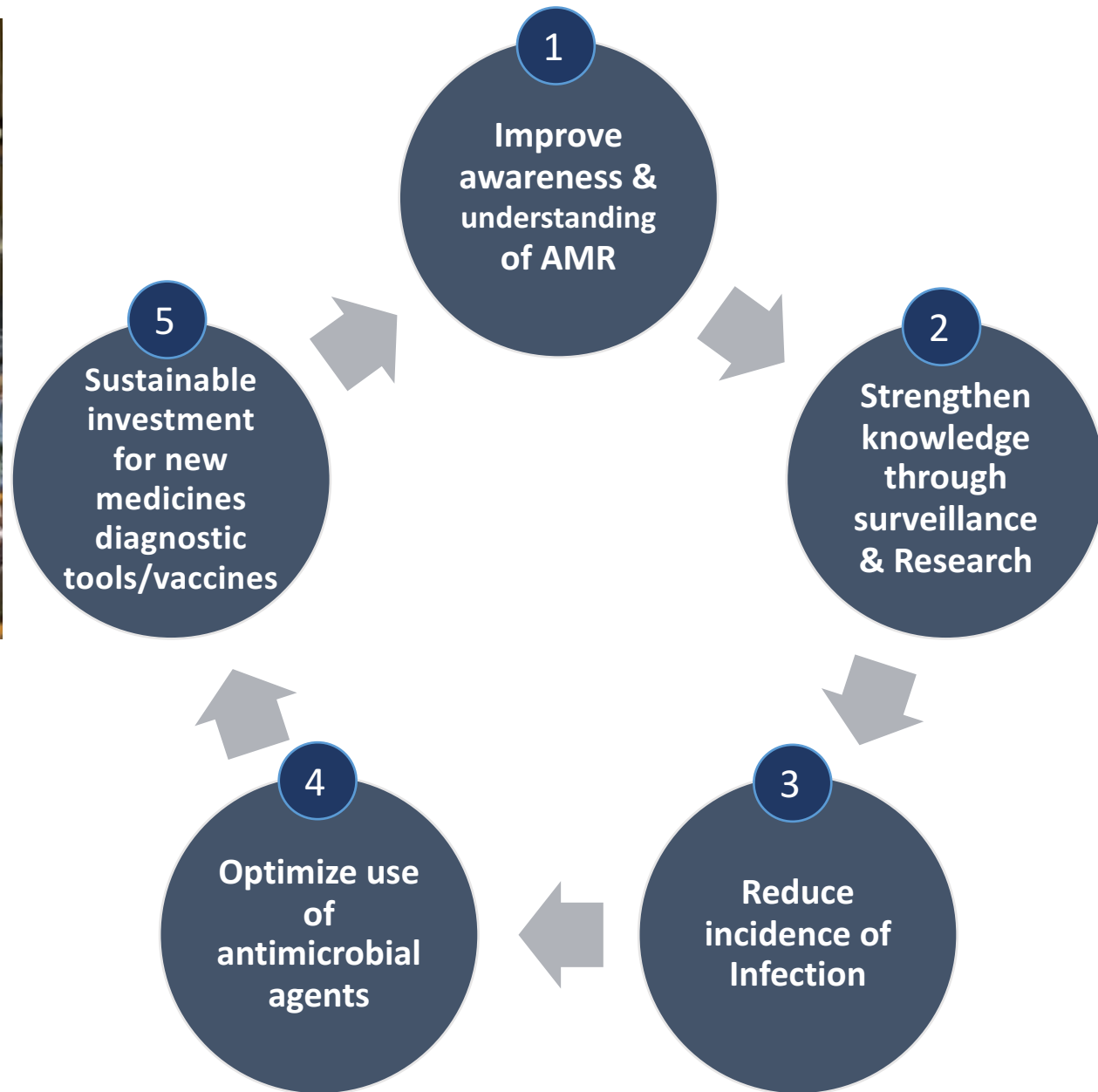
Timeline showing discovery of antibiotics, year of introduction and observation of resistance



- Discovery of antibiotics is rapidly followed by emergence of resistance against them.
- Continuing improvements are being made for discovery of new antimicrobial agents but are outpaced by emerging and re-emerging drug resistant infectious disease agents.



Sixty-eight World Health Assembly in May 2015 endorsed a global action plan to tackle antimicrobial resistance



Culture remains the gold standard for diagnosis of the Pathogen

So what is wrong?

- **As culture takes time – empirical based approach is used for diagnosis and prescription (broad-spectrum antibiotics are prescribed to cover as many pathogens)**
- **Leads to overtreatment or not the right treatment**
- **Impacts the emergence of antimicrobial resistance**
- **<60% sensitive; biological sample has low number of pathogens**
- **Complex etiology of conditions like sepsis makes it even more challenging - 90–95% of blood cultures remain negative in immunocompromised patients suspected of infection; even in the cases where bacterial or fungal sepsis is likely**



Sites of Infection

	No.	% Total
Lung	1016	37.2
Intraabdominal	801	29.3
Bowel perforation/peritonitis	226	8.3
Postoperative bowel perforation/anastomotic dehiscence	65	2.4
Spontaneous bacterial peritonitis	50	1.8
Other peritonitis	18	0.7
Intraabdominal abscess	44	1.6
Cholecystitis	40	1.5
Ascending cholangitis	43	1.6
Ischemic bowel/bowel infarction	166	6.1
<i>Clostridium difficile</i> enterocolitis/toxic megacolon	47	1.7
Genitourinary	293	10.7
Skin and soft tissue	197	7.2
Necrotizing soft tissue infections	74	2.7
Cellulitis	46	1.7
Operative wound infection	22	0.8
Soft tissue abscess	20	0.7
Decubitus ulcer	16	0.6
Diabetic lower extremity ulcer/cellulitis	13	0.5
Surgical site infection	31	1.1
Central nervous system infection (meningitis/abscess)	20	0.7
Intravascular catheter infection	100	3.7
Primary bloodstream infection (bacteremia without identifiable source)	120	4.4
Systemically disseminated infection (including yeast and tuberculosis)	58	2.1
Septic arthritis	21	0.8
Mediastinitis	15	0.5
Other	59	2.1

Sepsis is a systematic inflammatory response caused by infection

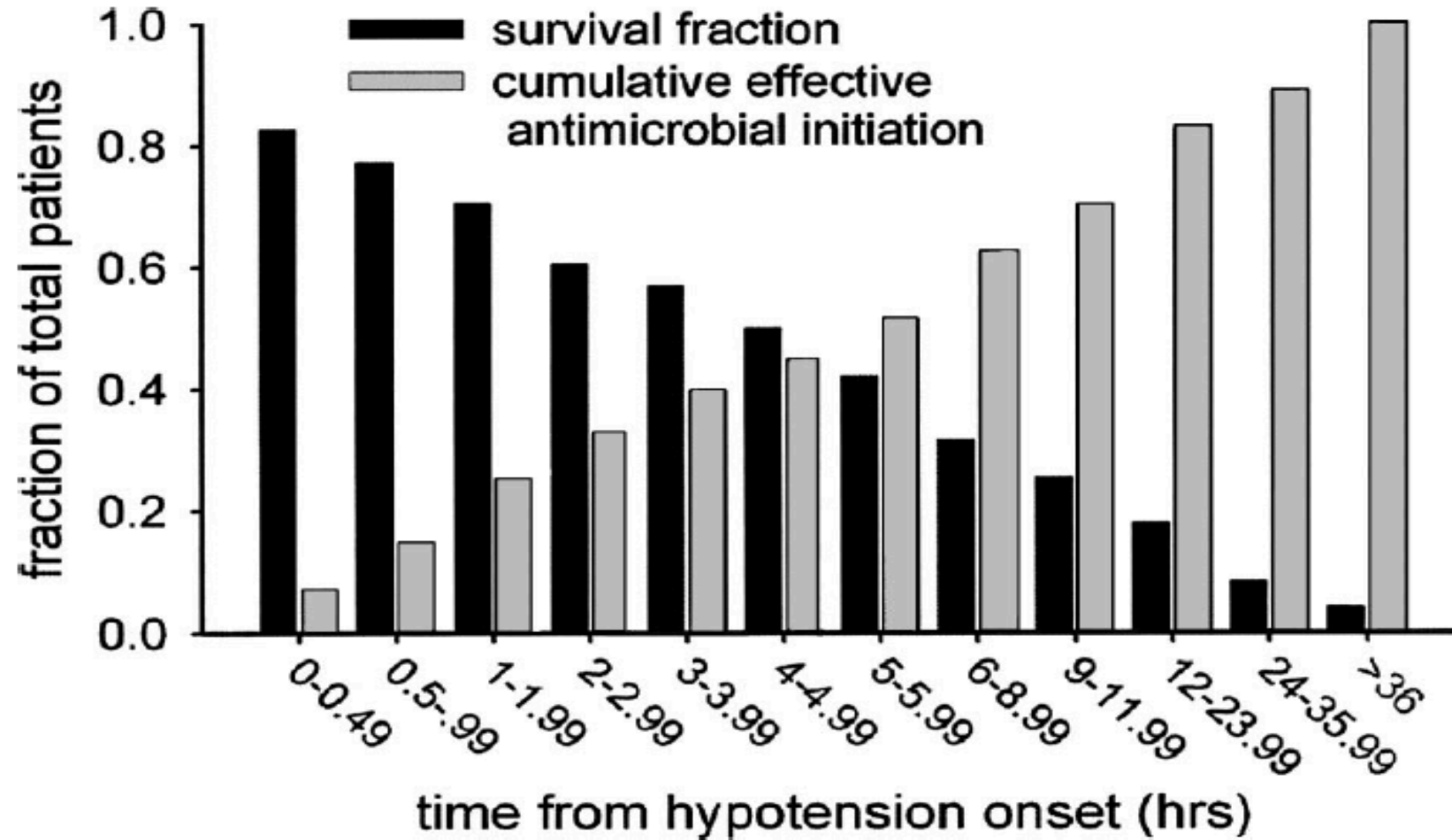
DeNIS Study



Pathogen	No. of Patients	% Total
Gram-negative organisms	930	47.9
<i>Escherichia coli</i>	435	22.4
<i>Klebsiella</i> species	131	6.7
<i>Pseudomonas aeruginosa</i>	115	5.9
<i>Enterobacter</i> species	80	4.1
<i>Haemophilus influenzae</i>	44	2.2
<i>Proteus</i> species	25	1.2
<i>Acinetobacter</i> species	21	1.1
<i>Serratia</i> species	20	1.0
<i>Stenotrophomonas maltophilia</i>	16	0.8
<i>Morganella morganii</i>	14	0.7
<i>Citrobacter</i> species	13	0.7
<i>Neisseria meningitidis</i>	6	0.3
<i>Burkholderia cepacia</i>	3	0.2
<i>Haemophilus parainfluenzae</i>	3	0.2
Other Gram-negative bacilli	8	0.4
Gram-positive organisms	731	38.3
<i>Staphylococcus aureus</i>	302	15.6
<i>Streptococcus pneumoniae</i>	170	8.8
<i>Streptococcus faecalis</i>	77	4.0
Group A <i>Streptococcus</i> species	69	3.6
Other β -hemolytic streptococci	43	2.2
<i>Viridans streptococci</i>	37	1.9
<i>Streptococcus faecium</i>	29	1.5
<i>Bacillus</i> species	5	0.3
<i>Corynebacterium jeikeium</i>	5	0.3
<i>Staphylococcus lugdunensis</i>	1	0.3
Yeast/fungi	160	8.2
<i>Candida albicans</i>	91	4.7
<i>Candida glabrata</i>	18	0.9
<i>Aspergillus/Mucor</i> species	14	0.7
<i>Blastomyces</i> species	10	0.5
<i>Candida tropicalis</i>	4	0.2
<i>Candida parapsilosis</i>	4	0.2
<i>Candida krusei</i>	3	0.2
<i>Cryptococcus neoformans</i>	1	0.1
<i>Histoplasma</i> species	1	0.1
Other unidentified yeast	13	0.6
Anaerobes	69	3.6
<i>Clostridium difficile</i>	46	2.4
<i>Bacteroides fragilis</i>	15	0.8
Other clostridia	8	0.4
<i>Legionella</i> species	8	0.4
<i>Mycobacterium tuberculosis</i>	11	0.6

Suspected primary pathogens in septic shock

The Need for Rapid Diagnosis



<https://pubmed.ncbi.nlm.nih.gov/16625125/>



Need a method that:

- Non culturable pathogen detection
- Can detect an entire range of pathogens
- Comprehensive
- Sensitive
- Specific
- Rapid
- Works with dead or live pathogen
- Differentiate between pathogen and human cells

Do Sequencing-based methods offer a solution?



What sequencing based methods can offer?

- Identify unculturable pathogens
- Distinguish between different pathogen strains
- Detect new drug resistance determinants, e.g. detection of *mecC* in *S. aureus**
- Rapid with extreme accuracy, -> better-informed treatment decisions
- Sequence data can be used to develop specific primers for PCR-based assays
 - to provide more complete sequence information and
 - to develop other specific field tests – specific genotype based

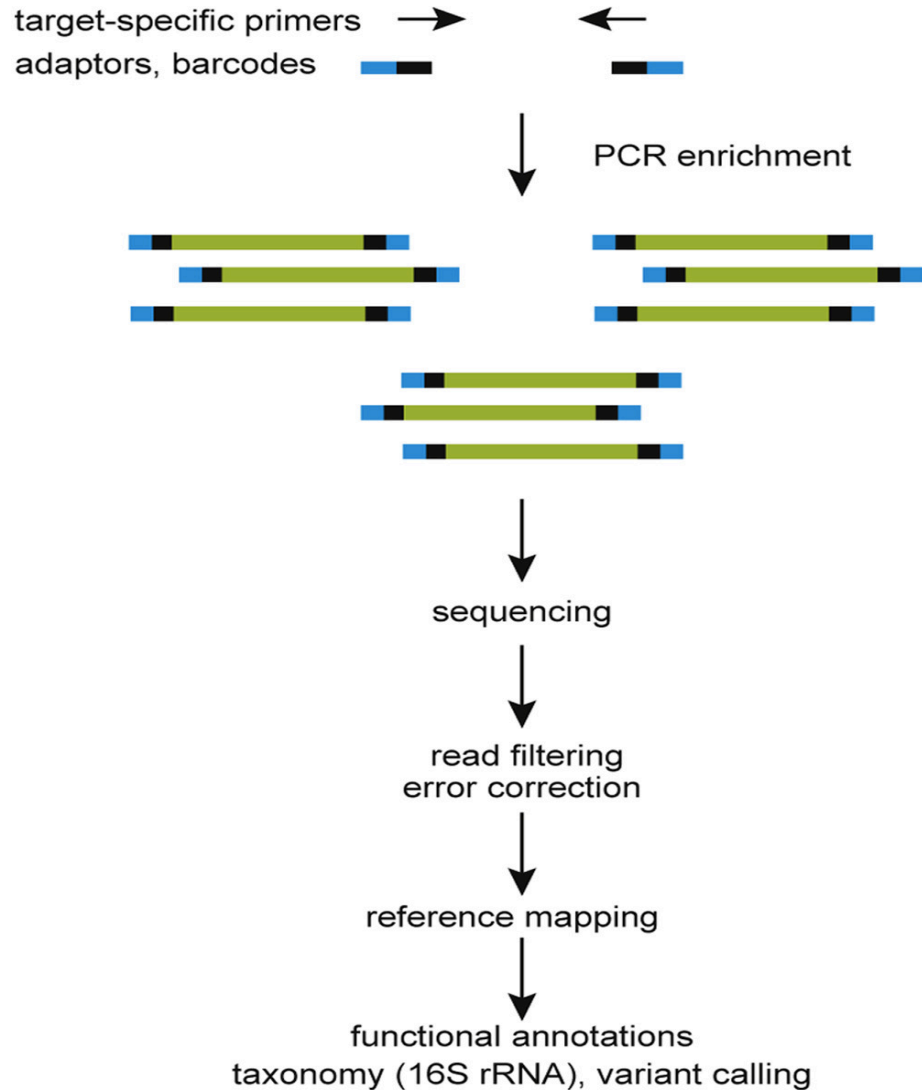
*<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3989053/>



Traditional vs Next Generation Sequencing Platforms in Clinical Microbiology

- The Sanger method has limited sensitivity for minor variants when present at <15% to 20% of the viral population, whereas NGS methods can detect drug-resistant mutations (DRMs) present at approximately 1%
- At least half of the DRMs identified by NGS are missed by Sanger sequencing
- The presence of such variants has been shown to predict an increased risk for therapy failure

Targeted amplicon sequencing

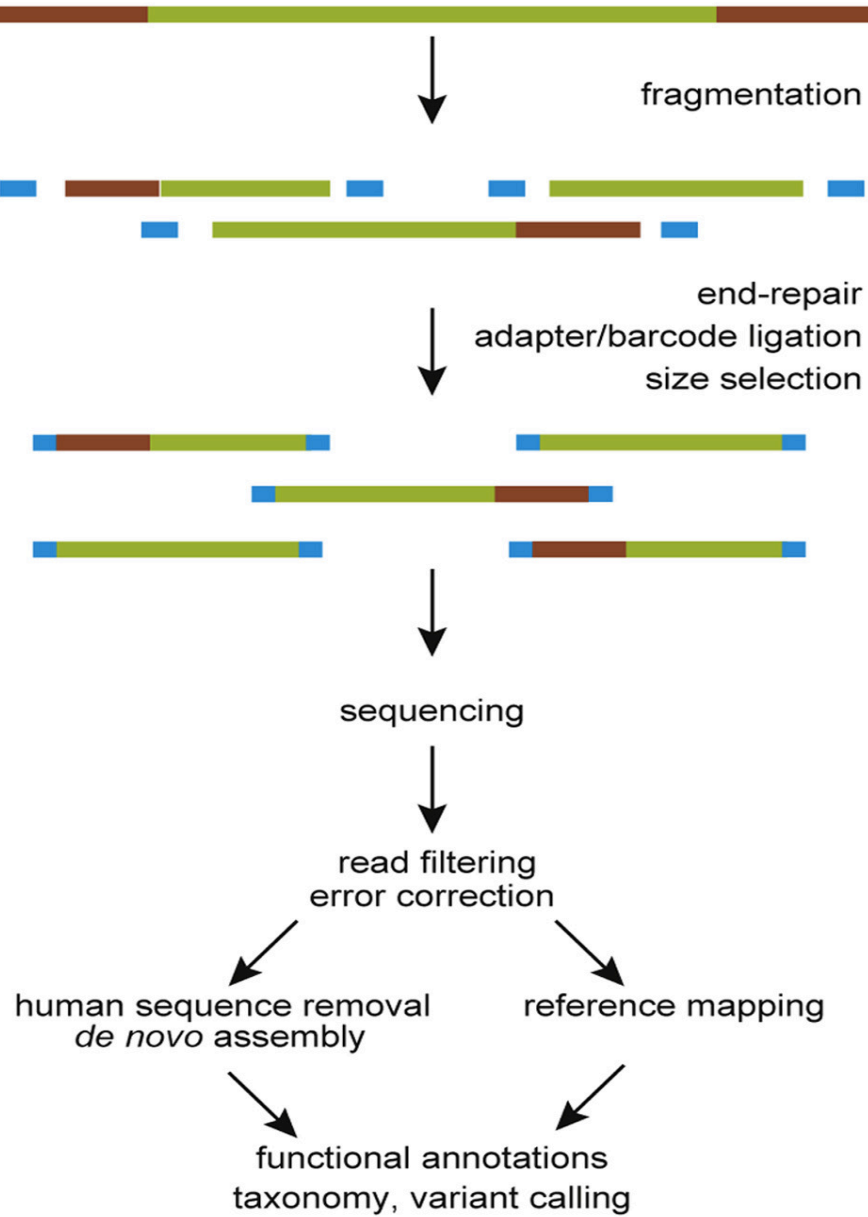


Cannot determine new pathogens or

New antimicrobial resistance genes

Not comprehensive

Whole-genome sequencing



Comprehensive

Can help in detecting novel AMR genes

Nanopore - Minlon



From bench to bedside – Point of care sequencing - promise

Companies which are pushing the boundaries of PoC with NGS in IDD

COMPANY	REGION	PRODUCT TYPE(S)	DESCRIPTION	FUNDING
Illumina and bioMérieux	U.S.	<ul style="list-style-type: none"> ▪ NGS Service 	Illumina, the dominant player in the NGS market, and bioMérieux, a leader in <i>in vitro</i> diagnostics, partnered together to launch an NGS service dedicated to infectious disease diagnostics. ¹⁴	<ul style="list-style-type: none"> ▪ Through a partnership between Illumina and bioMérieux, both global players in healthcare.¹⁵
Oxford Nanopore Technologies	U.K	<ul style="list-style-type: none"> ▪ NGS Sequencers ▪ NGS Assays ▪ Sample Prep Kits ▪ Library Prep Kits 	Founded in 2005, the company is developing portable NGS sequencing products for the analysis of single molecules. ^{22, 27}	<ul style="list-style-type: none"> ▪ Raised a total of \$386 million through VC funding. ▪ Closed a \$109 million round in July 2015 from new and existing investors in the U.K., U.S., and mainland Europe for product development, manufacturing and commercialization.²⁰
DNAe	U.K.	<ul style="list-style-type: none"> ▪ NGS Sequencers ▪ NGS Assays 	Founded in 2003, DNAe aims to revolutionize NGS diagnostic technology by bringing the entire NGS workflow from sample preparation, sequencing, and analysis into one semiconductor chip. ⁴	<ul style="list-style-type: none"> ▪ Secured a \$38 million bank facility from Citibank in November 2015 for development of an NGS <i>in vitro</i> diagnostic test for Serious Blood Infections.⁵ ▪ Completed a Series A fundraising round in April 2014 for an undisclosed amount.⁵
MRIGlobal	U.S.	<ul style="list-style-type: none"> ▪ NGS Sequencers ▪ NGS Assays 	Founded in 1977, MRIGlobal is an independent organization that performs contract research for the government and the industry.	<ul style="list-style-type: none"> ▪ Awarded \$14.8 million in February 2015 from the Defense Threat Reduction Agency of the U.S. Department of Defense to develop a comprehensive NGS platform for infectious diseases diagnosis.⁸
Pathoquest	France	<ul style="list-style-type: none"> ▪ NGS Assays ▪ Bioinformatics 	Pathoquest is developing a comprehensive NGS assay for diagnosing infectious diseases, including bioinformatics.	<ul style="list-style-type: none"> ▪ Raised \$5 million in July 2013 in Series B funding by IDInvest Partners, Aurinvest and Kurma Partners.²³

Species identification

From clonal sample

- NCBI BLAST
- GenBank
- Other genome databases in Table 1

From nonclonal sample

Meta-assembly

- AMOS
- MIRA
- MetaVelvet

Clustering and species annotation

- MEGAN
- MG-RAST

Maximum likelihood phylogeny trees

- BEAST
- RAxML
- ClonalFrame
- ClonalOrigin

Whole-genome alignment

For SNP calling

- Mummer
- Mugsy

For structural variant calling

- Mauve

Gene annotation

Bacterial

- GLIMMER
- RAST

Drug resistance in bacteria

- ResFinder
- ARG-ANNOT

A few databases and tools for DR determinant

In Infect Dis

Clin Infect Dis

Examples of Public Bioinformatics Databases That May Be Leveraged for Multiscale Analysis of Infectious Disease^a

Database Focus	For General Research	For Infectious Disease	
		Multipathogen	Pathogen-Specific
Genomes	<ul style="list-style-type: none"> • NCBI Nucleotide (GenBank/RefSeq) • ENA/EMBL • DDBJ 	<ul style="list-style-type: none"> • ViPR • NMPDR • PATRIC • EuPathDB 	<ul style="list-style-type: none"> • Influenza Research Database • Tuberculosis Database • LANL: Databases for HIV, HCV, and HFV
Gene products and functionality	<ul style="list-style-type: none"> • UniProt • KEGG 	<ul style="list-style-type: none"> • Pathogen-Host Interaction Database • Antibiotic Resistance Genes Database • Comprehensive Antibiotic Resistance Database 	
Expression and immune profiles	<ul style="list-style-type: none"> • GEO • ArrayExpress 	<ul style="list-style-type: none"> • ImmPort 	

List is not comprehensive



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Prediction of Susceptibility to First-Line Tuberculosis Drugs by DNA Sequencing

The CRyPTIC Consortium and the 100,000 Genomes Project

ABSTRACT

BACKGROUND

The World Health Organization recommends drug-susceptibility testing of *Mycobacterium tuberculosis* complex for all patients with tuberculosis to guide treatment deci-

The members of the writing group (Timothy M. Walker, D.Phil., A. Sarah Walker, Ph.D., and Tim E.A. White, D.Phil.) received



Lets know more about this study:

1. A total of 10,209 isolates were analyzed.
2. Resistance to isoniazid, rifampin, ethambutol, and pyrazinamide was correctly predicted with 97.1%, 97.5%, 94.6%, and 91.3% sensitivity, respectively
3. Susceptibility to these drugs was correctly predicted with 99.0%, 98.8%, 93.6%, and 96.8% specificity.
1. Among the 4037 phenotypic profiles that were predicted to be pansusceptible, 3952 (97.9%) were correctly predicted.

Susceptibility is predicted with higher accuracy

Clinical correlation of resistance phenotype is a limiting factor!!



Modes of action of first-and second-line anti-TB drugs, mechanisms of drug resistance and mutation frequency for each gene in clinical Mtb isolates

Group	Drug	Drug action	Drug resistance-associated gene(s)	Mutation frequency in clinical isolates (%) ^a
First-line anti-TB drugs	Rifampicin	Binding to the β -subunit of the RNA polymerase, inhibition of the elongation of messenger RNA	<i>rpoB</i> encoding for β -subunit of RNA polymerase	90–100
	Isoniazid	Activation by a catalase-peroxidase enzyme	<i>katG</i> encoding for catalase-peroxidase	40–97
		Inhibition of the synthesis of mycolic acids through binding to NADH-ACP-reductase	<i>inhA</i> encoding for fatty acid enoyl acyl carrier protein reductase A (InhA)	8–64
	Ethambutol	Inhibition of an arabinosyl transferase involved in cell wall synthesis	<i>embB</i> encoding for arabinosyl transferase	47–89
	Pyrazinamide	- Activation by the pyrazinamidase - Disruption of membrane energetics that inhibits membrane transport	<i>pncA</i> encoding for pyrazinamidase	44–97
	Streptomycin	Inhibition of protein synthesis by interaction with the 16S rRNA and the S12 ribosomal protein	<i>rrs</i> encoding for 16S rRNA subunit <i>rpsL</i> encoding for S12 ribosomal protein	12–26 40–68
		Inhibition of methylation of 16S rRNA	<i>gidB</i> encoding for 7-methylguanosine methyltransferase	5–13
Second-line anti-TB drugs	Amikacin, kanamycin, capreomycin	Inhibition of protein synthesis by interaction with the 16S rRNA	<i>rrs</i> encoding for 16S rRNA	40–90
	Kanamycin	Inhibition of acetyltransferase	<i>eis</i> encoding for aminoglycoside acetyltransferase	28–80
	Capreomycin	Inhibition of methylation of 16S rRNA & 23S rRNA	<i>tlyA</i> encoding for 2'-O-methyltransferase	4–13
	Ofloxacin, levofloxacin, moxifloxacin, gatifloxacin	Inhibition of the topoisomerase II (DNA gyrase) lead to the inhibition of DNA supercoiling	<i>gyrA</i> encoding for DNA gyrase subunit A and	70–90
			<i>gyrB</i> encoding for DNA gyrase subunit B	0–11
	Ethionamide	Inhibition of the synthesis of mycolic acids by interaction with NAD that inhibits the enoyl-ACP reductase	<i>inhA</i> encoding for fatty acid enoyl acyl carrier protein reductase A (InhA)	33–62
		Inhibition of metabolic activation by interaction with the transcriptional repressor of the Monooxygenase (EthA)	<i>ethA</i> encoding for EthA <i>ethR</i> encoding for transcriptional repressor EthR, NADH-ACP	46–72 0–4

In Greek, pan means 'whole'



[Proc Natl Acad Sci U S A](#). 2005 Sep 27; 102(39): 13950–13955.

PMCID: PMC1216834

Published online 2005 Sep 19. doi: [10.1073/pnas.0506758102](https://doi.org/10.1073/pnas.0506758102)

PMID: [16172379](https://pubmed.ncbi.nlm.nih.gov/16172379/)

Genetics

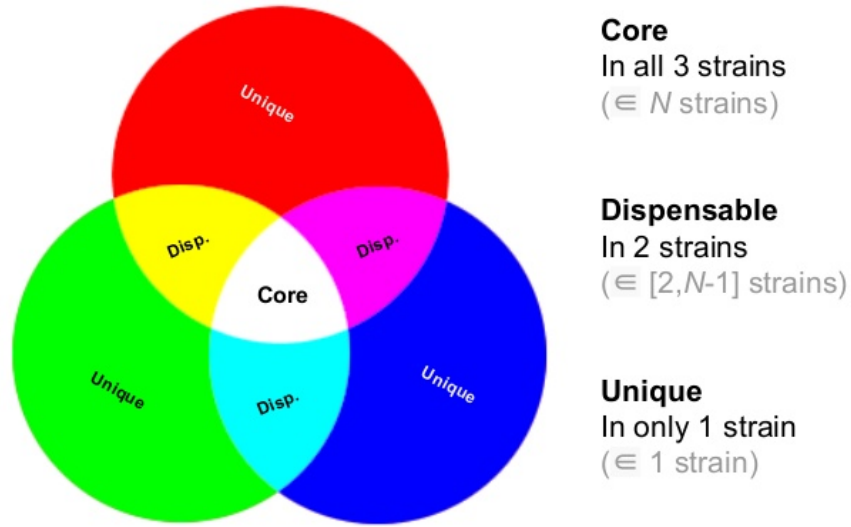
Genome analysis of multiple pathogenic isolates of *Streptococcus agalactiae*: Implications for the microbial “pan-genome”

[Hervé Tettelin](#)^{a,b}, [Vega Masignani](#)^{b,c}, [Michael J. Cieslewicz](#)^{b,d,e}, [Claudio Donati](#)^c, [Duccio Medini](#)^c,
[Naomi L. Ward](#)^{a,f}, [Samuel V. Angiuoli](#)^a, [Jonathan Crabtree](#)^a, [Amanda L. Jones](#)^g, [A. Scott Durkin](#)^a,
[Robert T. DeBoy](#)^a, [Tanja M. Davidsen](#)^a, [Marirosa Mora](#)^c, [Maria Scarselli](#)^c, [Immaculada Margarit y Ros](#)^c,
[Jeremy D. Peterson](#)^a, [Christopher R. Hauser](#)^a, [Jaideep P. Sundaram](#)^a, [William C. Nelson](#)^a, [Ramana Madupu](#)^a,
[Lauren M. Brinkac](#)^a, [Robert J. Dodson](#)^a, [Mary J. Rosovitz](#)^a, [Steven A. Sullivan](#)^a, [Sean C. Daugherty](#)^a,
[Daniel H. Haft](#)^a, [Jeremy Selengut](#)^a, [Michelle L. Gwinn](#)^a, [Liwei Zhou](#)^a, [Nikhat Zafar](#)^a, [Hoda Khouri](#)^a, [Diana Radune](#)^a,
[George Dimitrov](#)^a, [Kisha Watkins](#)^a, [Kevin J. B. O'Connor](#)^h, [Shannon Smith](#)ⁱ, [Teresa R. Utterback](#)ⁱ, [Owen White](#)^a,
[Craig E. Rubens](#)^g, [Guido Grandi](#)^c, [Lawrence C. Madoff](#)^{e,j}, [Dennis L. Kasper](#)^{e,j}, [John L. Telford](#)^c,
[Michael R. Wessels](#)^{d,e}, [Rino Rappuoli](#)^{c,k,l} and [Claire M. Fraser](#)^{a,b,k,m}

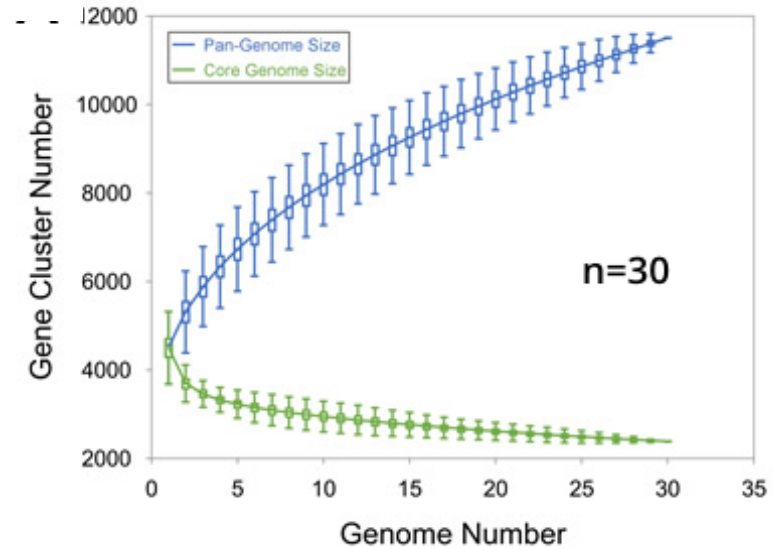
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Strains of a bacterial species might differ substantially in their gene content and total gene pool of a species might be orders of magnitudes larger than the gene content of any single strain

Three genomes



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The core genes are responsible for basic aspects of the biology of the species and its major phenotypic traits;

The accessory genes and singletons usually pertain to supplementary biochemical pathways and functions that may confer selective advantages such as ecological adaptation, virulence mechanisms, antibiotic resistance, or colonization of a new host.

Article | [OPEN](#) | Published: 17 October 2018

Machine learning and structural analysis of *Mycobacterium tuberculosis* pan-genome identifies genetic signatures of antibiotic resistance

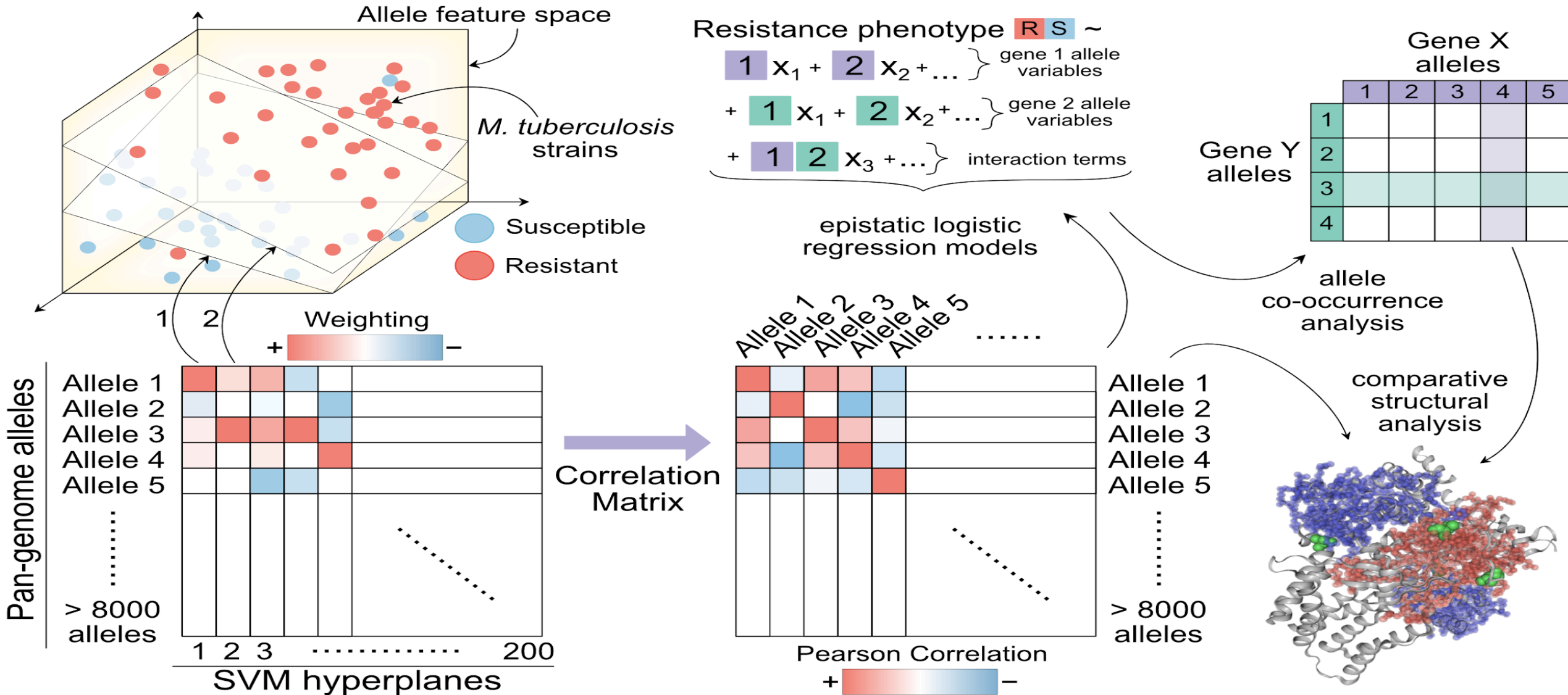
Erol S. Kavvas, Edward Catoi, Nathan Mih, James T. Yurkovich, Yara Seif, Nicholas Dillon, David Heckmann, Amitesh Anand, Laurence Yang, Victor Nizet, Jonathan M. Monk✉ & Bernhard O. Palsson✉

Nature Communications **9**, Article number: 4306 (2018) | [Download Citation](#)↓

The dataset – Genome IDs with Drug Sensitivity Profiling

	genome_id	isoniazid	rifampicin	ethambutol	amikacin	ciprofloxacin	clofazimine	cycloserine	ethionamide	kanamycin	moxifloxacin	nicotinamide
1	1295764_3	R	R	R			S		R	R		R
2	1423468_3	R	R	S								
3	1448744_3	R	R	S								
4	1448649_3	S	S	S								
5	1773_371	R	R	R								
6	1421929_3	R	R									
7	1448748_3	S	S	S								
8	1455273_3	R	R	R								
9	1455275_3	R	R	R								
10	1427320_3	R	R	R	R			R	R	R		
11	1427186_3	R	R	S	S			S	S	S		
12	1448825_3	S	S	S								
13	1447514_3	R	R	S								
14	1408948_4	S	R	S	S			S	S	S	S	
15	1421939_3	R	R									
16	1447458_3	R	S	S								
17	1773_369	S	S	S								
18	1773_368	S	S	S								
19	1773_365	R	R	R	R			S				
20												

Illustration of multi-layered analysis workflow



Known AMR genes uncovered by machine learning

Antibiotics	Known AMR genes
Isoniazid	<i>katG</i> ⁴³ , <i>inhA</i> ^{a20} , <i>fabG1</i> ⁴⁴
Rifampicin	<i>rpoB</i> ⁴⁵ , <i>rpoC</i> ^{a46} , <i>Rv3239c</i> ⁴⁷
Ethambutol	<i>embB</i> ⁴⁸ , <i>embC</i> ¹⁷ , <i>ubiA</i> ^{a6} , <i>embR</i> ^{a17}
Pyrazinamide	<i>pncA</i> ⁴⁹
Streptomycin	<i>rpsL</i> ⁵⁰ , <i>gidB</i> ⁵¹
Ofloxacin	<i>gyrA</i> ⁵²
4-Aminosalicylic acid	<i>folC</i> ^{a7} , <i>thyA</i> ^{a53}
Ethionamide	<i>ethA</i> ⁵⁴ , <i>inhA</i> ^{a20}
Known AMR genes associated with other antibiotics	<i>dprE1</i> ⁵⁵ , <i>ald</i> ⁵ , <i>alr</i> ⁵⁶ , <i>murA</i> ⁵⁷ , <i>pks2</i> ⁵⁸ , <i>pks12</i> ⁵⁹ , <i>ppsA</i> ⁶⁰ , <i>ppsD</i> ⁶⁰ , <i>drrB</i> ⁶¹ , <i>drrC</i> ⁶¹ , <i>moeW</i> ⁵⁵ , <i>Rv0687</i> ⁶² , <i>mshD</i> ⁶³ , <i>gyrB</i> ⁵² , <i>Rv1877</i> ⁶⁴ , <i>Rv0194</i> ⁶⁵

Gene	Drug
<i>Rv3848</i>	EMB, XDR
<i>embR</i>	EMB
<i>Rv3129</i>	EMB
<i>proC</i>	EMB
<i>kdpC</i>	EMB
<i>oxcA</i>	INH
<i>chp2</i>	ETA
<i>lipD</i>	ETA
<i>Rv3471c</i>	ETA, XDR, SM
<i>mmpL11</i>	PAS
<i>Rv0044c</i>	PAS
<i>Rv0954</i>	PAS
<i>Rv2560</i>	PZA
<i>Rv2090</i>	RIF, INH
<i>lpqZ</i>	RIF
<i>Rv1597</i>	RIF, MDR, INH
<i>Rv1543</i>	RIF, MDR
<i>nuoL</i>	MDR, PAS
<i>dnaA</i>	SM
<i>yajC</i>	SM
<i>accD5</i>	OFX, MDR
<i>Rv3041c</i>	RIF, OFX, SM, MDR
<i>VapC21</i>	XDR

Newly proposed AMR genes

README.md

KOVER_{2.0}

DOI [10.5281/zenodo.2630879](https://doi.org/10.5281/zenodo.2630879) build passing

Kover is an *out-of-core* implementation of rule-based machine learning algorithms that has been tailored for genomic biomarker discovery. It produces highly interpretable models, based on k-mers, that explicitly highlight genotype-to-phenotype associations.

Introduction

Understanding the relationship between the genome of a cell and its phenotype is a central problem in precision medicine. Nonetheless, genotype-to-phenotype prediction comes with great challenges for machine learning algorithms that limit their use in this setting. The high dimensionality of the data tends to hinder generalization and challenges the scalability of most learning algorithms. Additionally, most algorithms produce models that are complex and difficult to interpret. We alleviate these limitations by proposing strong performance guarantees, based on sample compression theory, for rule-based learning algorithms that produce highly interpretable models. We show that these guarantees can be leveraged to accelerate learning and improve model interpretability. Our approach

2.0.3 - Squirrel Monkey Latest

on Apr 6, 2019

+ 10 releases

Contributors 2

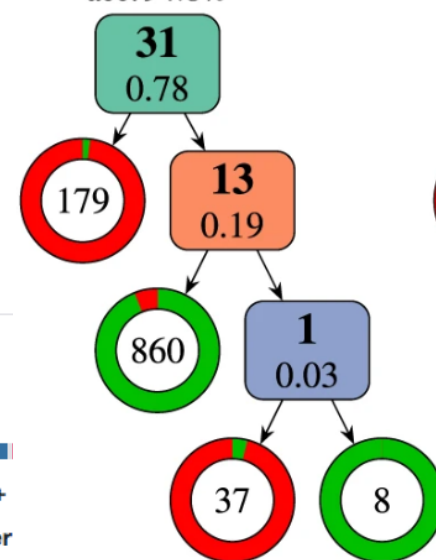
 aldro61 aldro61

 gletarte gletarte

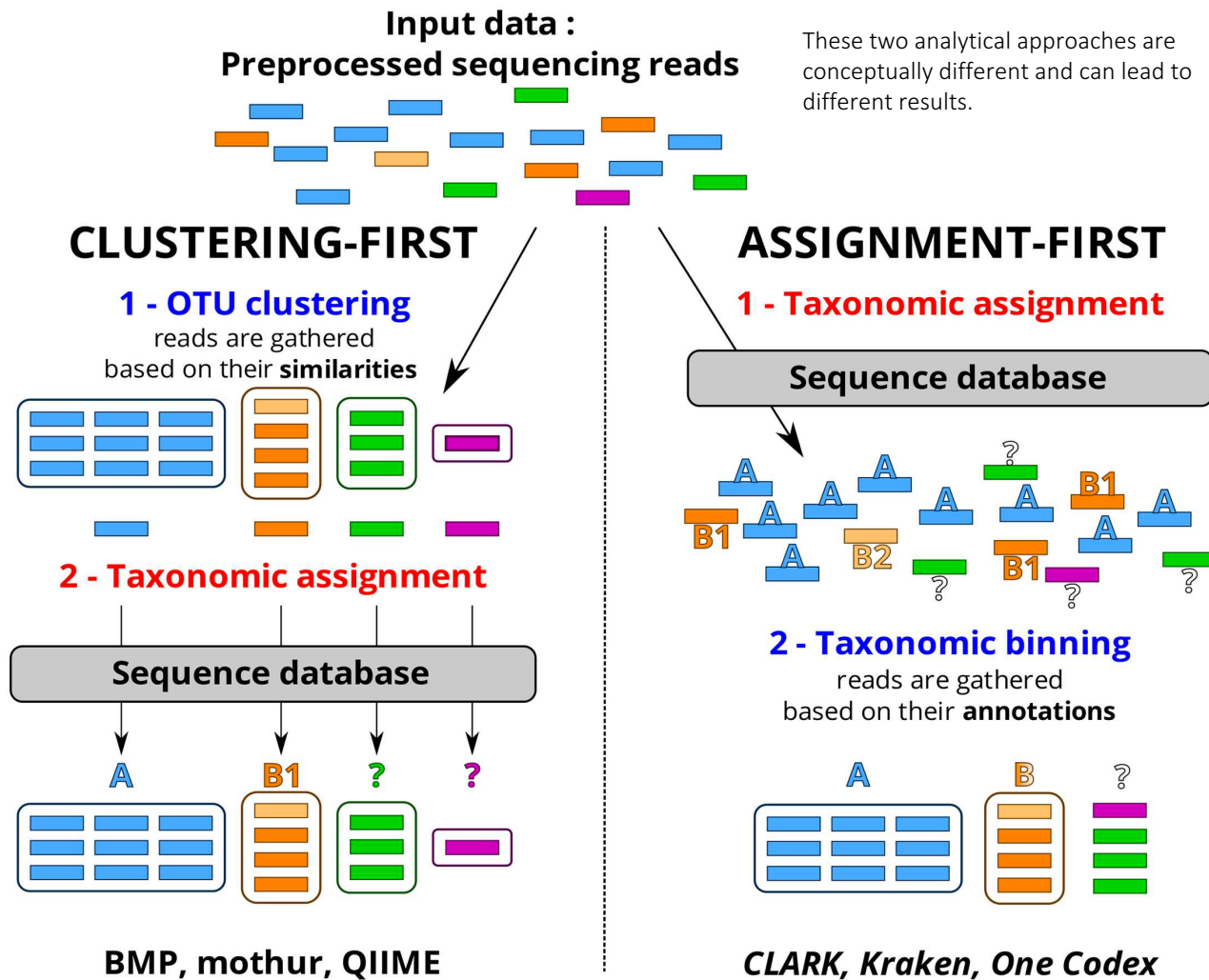
Languages

Python 71.5% C++
CMake 1.6% Other

CART_b
acc: 94.8%



Identification of strains from metagenome sequencing



mothur does not include taxonomic annotations at the species level.

For the QIIME integrated databases, less than 7% of Greengenes sequences and less than 45% of SILVA sequences are annotated at the species level.

KrakenUniq - NCBI Taxonomy IDs - hundreds of strains share the same taxonomy ID



~83K complete microbial genomes, including 50K distinct bacterial genomes, 27K viral genomes, and hundreds of archaeal and eukaryotic genomes.

- **Clinical metagenomics (CMg)** refers to the concept of sequencing the DNA of a clinical sample (without any prior culturing step) with the purpose of recovering clinical information
- **“Salvage microbiology”** is a term used for the application of molecular diagnostic techniques in the detection of bacterial DNA directly from clinical specimens
- Enable the detection of pathogens and of some antibiotic resistance genes (ARGs)

Knowledge of the full drug susceptibility profile would enable tailored treatment to improve efficacy and reduce exposure to ineffective toxic drugs



Details

File Properties

Owner
Andy Davies

Uploader
Andy Davies

Created
Apr 29, 2019, 10:07 PM

Modified
Apr 29, 2019, 10:07 PM

Size
24.2 KB

WIMP methods

Useful things to know:

- Reads can only be assigned to organisms that are included in the database and taxonomic subtree e.g. reads will not be assigned as dog (*Canis canis familiaris*) as this is not included in the database however, many dog reads will likely be classified as "mammalia" as they will have homology to the human reference which is included in the database.
- The WIMP application is optimised for Oxford Nanopore Technologies DNA sequence reads (both length and error profile)
- The application works best with genomic DNA sequences from whole genome or metagenome library preparation. Amplicon sequences such as the 16S rRNA sequence do not contain sufficient taxonomic context for this WIMP method to perform accurate sequence classification.
- The normalised qscore presented in the results is calibrated against the Zymo microbial mock community collection.

Generating WIMP database:

References / data source

WIMP classifies DNA sequence reads against a pre-generated [centrifuge](#) index. This index has been prepared from a DNA sequence collection that includes both the RefSeq microbial genome reference sequences and the primary chromosome assemblies from the human genome ¹. The reference genome sequences are downloaded from the NCBI primary sequence database. [Dustmasker](#) is used to filter sequence regions of low complexity, in addition to this we mask adapter sequences. The adapters masked are provided [here](#). The WIMP database is generated using the `centrifuge-build` script and is run on an amazon x1e.2xlarge instance. The `centrifuge-inspect` method is used to collate the information for the construction of the taxonomic subtree for the genomic sequences included in the centrifuge database.

Database details:



DNA sequencing as the new microscope

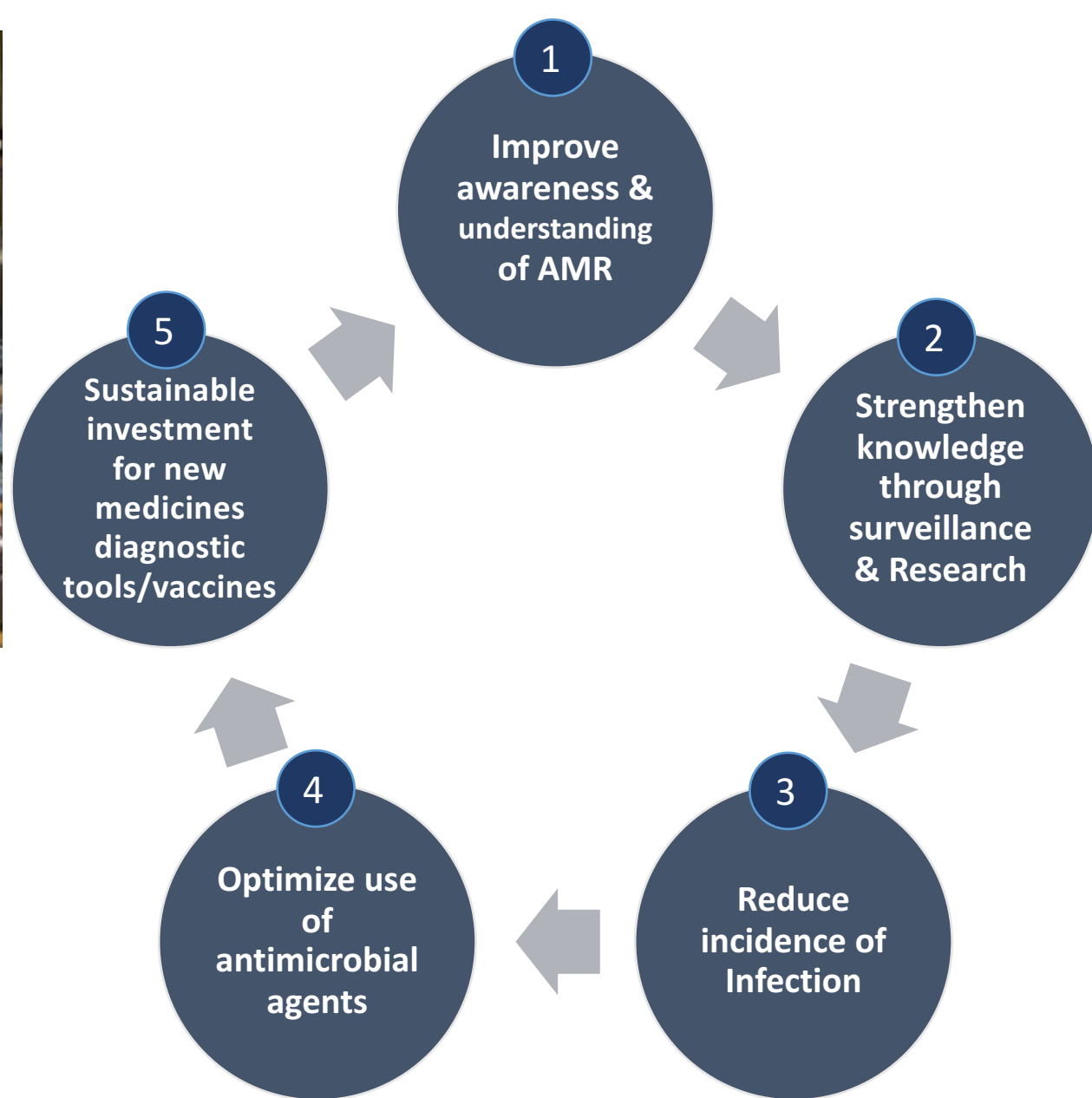
- It has been about **400 years** since the invention of **light microscopy**, a technology which continues to be used and to evolve.
- It has been only **40 years** since the invention of **DNA sequencing**; the technologies and likely to also continue to develop in the coming decades and centuries.
- It has transformed biomedical research, and **is beginning to transform clinical medicine**
- DNA sequencing will have a **longevity and impact on par with or exceeding that of the microscope.**

doi:10.1038/nature24286





Sixty-eighth World Health Assembly in May 2015 endorsed a global action plan to tackle antimicrobial resistance

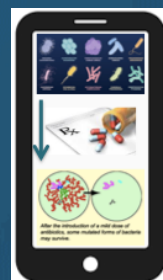


Open Antimicrobial Resistance Awareness Drive

Abot – knowledge repository for education & awareness on AMR



- Bot learning
- Structured knowledgebase
- Outreach



A Chatbot will guide the Players on the most viable treatment options

Basic – Treat drug susceptible cases
Advanced – Treat drug resistance cases
Faster cure is the winning bet

A Games on education & awareness on AMR



- Game Designer / creative skills
- Gamers
- Coders

<https://freshdesk.com/help-desk-software/chatbot-conversations-dos-donts-blog/>



CSIR-IMTECH



SEARCH

RAPHAEL



OPEN AMR AWARENESS DRIVE

Build a community of Gamers for creating awareness on antimicrobial resistance

ADDED ON THE 2 JUN 2019

UPDATED ON THE 17 SEP 2019

BIOLOGY

GAME

DIGITAL

EDUCATION

Aryabot Learning about AMR



Good evening



I'm Arya! I'm here to help you know about Antimicrobial resistance (AMR) and its effects.

I'm designed to create awareness about AMR.



Before we get started, could you please introduce yourself

Anshu



Hi Anshu! Let's start by a question: do you know what superbugs are?

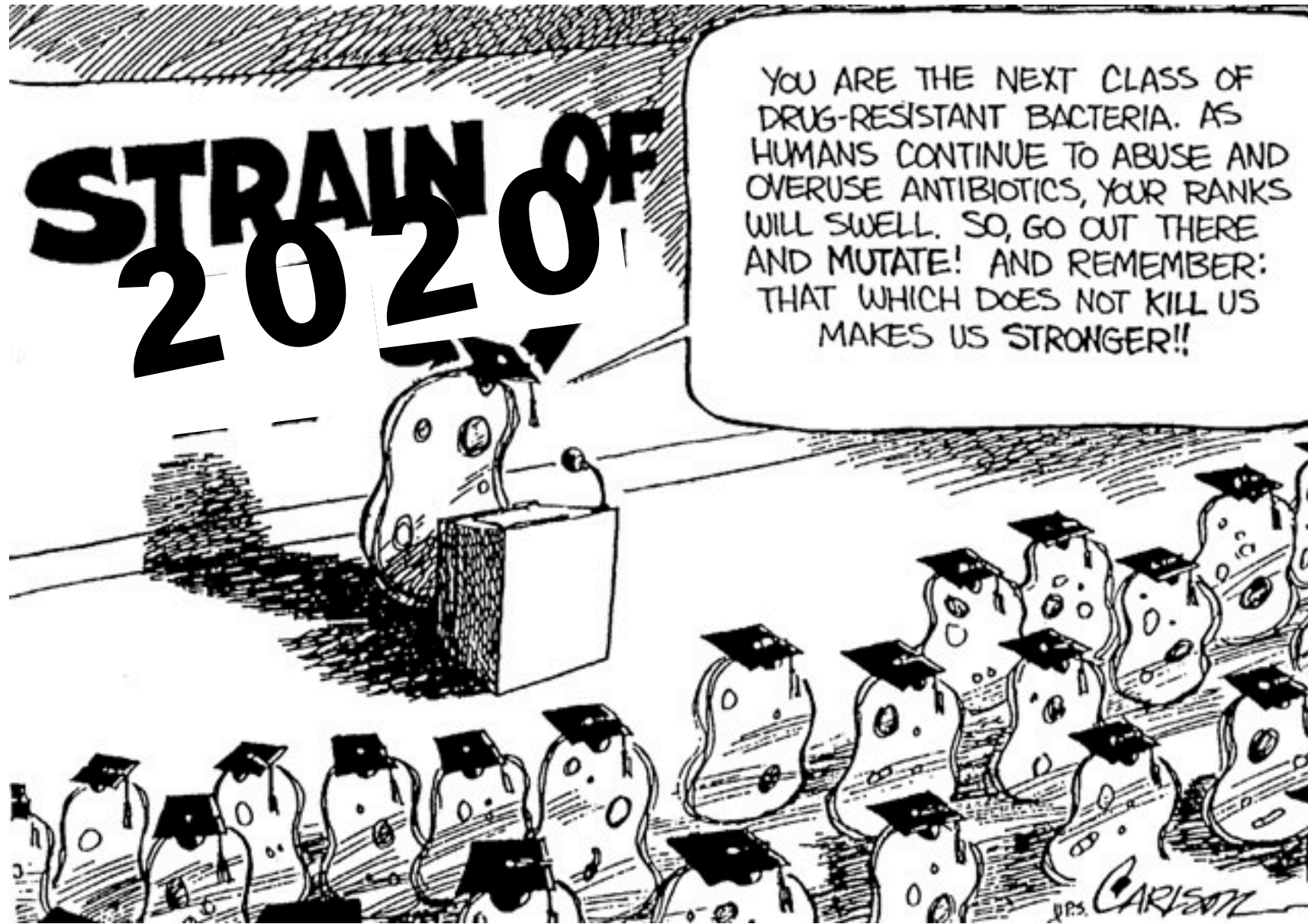
Type a message...



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Playtest and Feedback Sessions with Aryabot



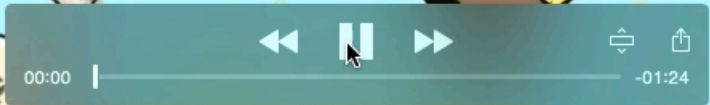


**How do we
convey the
same in a
video game?**

<https://slideplayer.com/slide/4685976/>



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Antibiotic Categorization

To address these issues, WHO developed a framework based on three different categories – Access, Watch and Reserve – which all together forms the AWaRe categorization of antibiotics.

ACCESS GROUP

- first or second choice antibiotics
- offer the best therapeutic value, while minimizing the potential for resistance

WATCH GROUP

- first or second choice antibiotics
- only indicated for specific, limited number of infective syndromes
- more prone to be a target of antibiotic resistance and thus prioritized as targets of stewardship programs and monitoring

RESERVE GROUP

- "last resort"
- highly selected patients (life-threatening infections due to multi-drug resistant bacteria)
- closely monitored and prioritized as targets of stewardship programs to ensure their continued effectiveness

Game specifics are based on AWaRe

Initial game levels cannot unlock antibiotics in Watch / Reserve categories

In order to unlock these, the players should accumulate points in earlier layers by assigning right antibiotic for the pathogens

Community Collaborative Projects – Crowdsourcing @ work



SysBorg 2.0

PMID: 21782516

OSDD Chem



PMID: 22808064
PMID: 23629487
PMID: 25304862

BioPhytMol

PMID: 25360160

OSDDChemDesign

Cloning, Expression
and Purification of
Proteins



dPABBs
Design Peptides Against Bacterial Biofilms
PMID: 26912180

Drug Repurposing through Network-Based Inference

Systems level modeling for identification of drug targets for gram negatives

doi.org/10.1007/s12039-017-1268-4

MPDS^{TB}

RepTB

PMID: 29785561



2009

2010

2011

2012

2013

2014

2015

2016

2017----2019/20



PMID: 19758471

E-MiDAS



PMID: 23585830

SSCUE
PMID: 24434286



PMID: 26244889

Identification of Clinically actionable variants in case of mt diseases – applicable to complex diseases

PMID: 26180633

MitoLINK
Community Collaborative Systems Level Platform



CSIR-IMTECH

Worked with over 500 students on several genomics and drug discovery projects





dPABBs: A Novel *in silico* Approach for Predicting and Designing Anti-biofilm Peptides

SCIENTIFIC REPORTS

OPEN

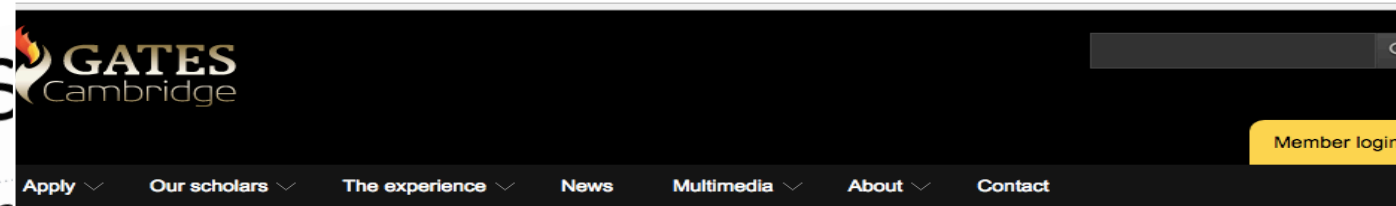
dPABBs: A Novel *in silico* Approach for Predicting and Designing Anti-biofilm Peptides

Arun Sharma^{1,2,*}, Pooja Gupta^{1,3,*}, Rakesh Kumar^{1,2} & Anshu Bhardwaj^{1,2}

Received: 03 July 2015
Accepted: 27 January 2016
Published: 25 February 2016

Increasingly, biofilms are being recognised for their causative role in persistent infections (like cystic fibrosis, otitis media, diabetic foot ulcers) and nosocomial diseases (biofilm-infected vascular catheters, implants and prosthetics). Given the clinical relevance of biofilms and their recalcitrance to conventional antibiotics, it is imperative that alternative therapeutics are proactively sought. We have developed dPABBs, a web server that facilitates the prediction and design of anti-biofilm peptides. The six dPABBs models implemented on dPABBs were observed to identify anti-biofilm peptides on 100% accuracy and Weka models implemented on dPABBs were observed to identify anti-biofilm peptides on 100% accuracy. On the N-terminus, it was seen that either of the cationic residues (maximum accuracy, sensitivity, specificity and MCC of 95.24%, 92.50%, 97.73% and 97.73% respectively, on the training datasets). On the C-terminus, it was seen that either of the cationic residues, R and K, is present at all five positions in case of the anti-biofilm peptides. On the other hand, the uncharged polar residue S is preponderant at the first (also anionic polar residue D) and fifth positions. Positive predictions were also obtained for 29 FDA-approved peptides and ten antimicrobial peptides in clinical development, indicating at their possible repurposing for biofilm therapy. dPABBs is freely accessible on: <http://ab-openlab.csir.res.in/abp/antibiofilm>

<https://www.gatescambridge.org/members-area/connect/directory/scholar/14672>



Home / Members' area / Connect / Directory / Scholar

Profile

Go to [find a scholar](#)

Miss Pooja Gupta (2018)

Scholar-elect
India
MPhil Biological Science at the Department of Biochemistry
Sidney Sussex College



Biography

I was born and raised in New Delhi and am currently completing my undergraduate studies in Microbiology at the University of Delhi. Growing up on a steady diet of science fiction and popular science, I was certain by the time I was 17 that I wanted to pursue a career in research, and my journey which began under Dr. Anshu Bhardwaj's guidance at the Council of Scientific and Industrial Research (CSIR), India, has been quite serendipitous since. I am keenly interested in the structure-guided fragment-based method being employed in Prof. Sir Tom Blundell's lab for antimycobacterial drug discovery. Using this method, one can not only design highly selective ligands against validated targets, but perhaps also implement the ambitious multidrug and polypharmacological strategies for more efficacious drug regimens with fewer side-effects. I hope to better understand aspects of bacterial community behaviour and explore how it may be exploited for more specific antimicrobial therapy. Learning techniques in Structural Biochemistry, principles of Drug Design, Systems-level approaches and Synthetic Biology could eventually enable an integration of these fields to study interactions in microbial communities, along with emergent phenomena like biofilm formation and drug resistance. I would also like to teach students and dedicate time to science communication for public engagement, which I believe is crucial to the larger scientific cause. It would be an exciting challenge to try and capture popular imagination with advanced

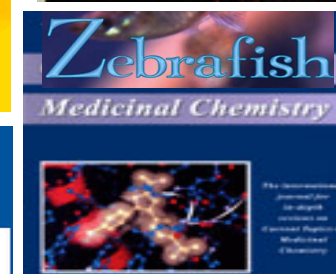


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Individual Competence to Team Delivery



And more...



AB-OpenLab – India & France



- OSDD Community
- JOGL
- LWB

anshu@imtech.res.in

anshu.bhardwaj

[@anshub](https://twitter.com/anshub)



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